

List Products and Services

IVD GmbH

Analytical Laboratory accredited by the German Accreditation Authority (DAkks)
according to the European standard DIN EN ISO/IEC 17025

Farm Animals: SWINE, RUMINANTS & POULTRY

HUMAN



IVD GmbH Innovative Veterinary Diagnostics
Albert-Einstein-Str. 5
30926 Seelze-Letter
Germany



for *Leptospira* spp.



Version of March 2025

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General information

Samples are analysed individually. All prices listed are per sample and do not include statutory VAT. Please note that we do not accept payment via cheque.

Our General Terms and Conditions of Sale apply to all services; these can be viewed in our premises or on our homepage: <https://www.ivd-gmbh.de>.

Our privacy policy can be found at <https://www.ivd-gmbh.de/datenschutz>.

Please contact us in case of special diagnostic services, analyses or questions. We are prepared to answer any question and consider any request within the areas of our professional and technical expertise.

There is a surcharge for diagnostic services below a minimum charge of € 10.00.

A surcharge plus VAT will be assessed for retroactive modifications of orders authorised/requested/required by the customer.

Selection and submission of samples

The materials best suited for serological examinations are serum or blood without anticoagulants.

The use of plasma (addition of EDTA, Li-heparin, citrate) can cause artefacts in the complement fixation test (CFT) and in agglutination reactions (HI, MAT, RBT and others).

Tissue and swab samples collected immediately *post mortem* from typical lesions are often more suitable for the molecular biological and cultural detection of agents than samples which can be taken *intra vitam*.

Shipping recommendations

When pathogen levels are low, both analytical sensitivity and diagnostic certainty of molecular biological analyses are enhanced by immediate shipment of cooled samples, due to retardation of autolysis. Chill fresh tissues prior to shipment and include frozen gelpacks with the shipment; enclose paperwork in waterproof packaging. When shipping fresh tissues, consider the possibility of shipping delays due to weekends or holidays.

Information on regulations for the shipment of hazardous materials can be found on our home page: <https://www.ivd-gmbh.de>.

The IVD GmbH laboratory is accredited according to European standard DIN EN ISO /IEC 17025. IVD has performed diagnostic testing for infectious agents of farm animals, horses, dogs, cats and exotic animals since 1997.

Tests that do not conform to the regulations of the national accreditation body of Germany are marked with a superscript circle (°).

In test reports, only the method is reported, when using commercial test kits also the manufacturer is mentioned, not the batch number of the test kit or the version number of the manual (simplified report).

Your personal contact**Direct dial**

| | | |
|---|--------------------------|-----------|
| Central Office | +49 (0)511-220029 | -0 |
| Managing Partners | | |
| Katrin Strutzberg-Minder MSc, DSc | | -0 |
| Matthias Homuth DVM, Specialist in Veterinary Microbiology | | -0 |
| Jens-Peter Minder Industrial Manager | | -0 |
| Quality Management | | |
| Astrid Ullerich DVM | | -12 |
| Serology / Parasitology | | |
| Sebastian Fischer DVM, Specialist in Veterinary Microbiology | | -22 |
| Molecular Biology | | |
| Jan Böhmer DVM | | -30 |
| Bacteriology | | |
| Mira Schumann BSc | | -40 |
| Ina Zerbin DVM | | -65 |
| Pathology | | |
| Maren Biesler DVM | | -13 |
| Administration | | |
| Centrale office | | -0 |
| Accounting | | -84 |

MATERIAL FOR SAMPLING, PACKAGING AND TRANSMITTAL OF SPECIMEN

The IVD GmbH provides its customers with material for taking samples of meat juice, oral fluid as well as sample containers for histological examination in smaller packaging units. The packaging corresponds to ADR regulation P650 for mail delivery to our laboratory.

| | Minimum purchase/ packaging unit (PU) | Price in € per PU |
|---|--|----------------------|
| Suesse Post Box (suitable for 12-16 blood samples or 5 fecal samples) incl. 1 protective bag with absorbent insert | 20 pc. | 23.70 |
| Suesse Post Box Maxi (suitable for 24-32 blood samples or 10 fecal samples) incl. 2 protective bags with absorbent insert | 15 pc. | 26.20 |
| Protective Bag with absorbent insert (for 6-8 blood or 5 fecal samples) | 50 pc. | 20.20 |
| Protective Bag LARGE with absorbent insert (for 3 fecal poaches) | 25 pc. | 14.90 |
| Protective Container with absorbent insert and screw cap (only for single samples) | 50 pc. | 16.10 |
| Fecal Tube with screw cap 76x20 mm | 100 pc. | 17.80 |
| Formalin filled sample tubes for histological examinations (100 ml urine cup with screw cap filled with 50 ml 4% Formalin) | 40 pc. | 17.80 |
| Meat Juice Collector with Stopper | 70 pc. | 16.10 |
| FTA Cards (4 spots each) | 1 pc. | 6.50 |
| DBS (Dried Blood Spots - 5 spots each) | 5 pc. | 10.10 |
| Cold Pack (according to availability) | max. 1 pc. per box | free of charge |
| Mycoplasma Medium | on request | |
| Fecal Sampling Horse & Small Animal Kit each shipping box (Suesse Post Box) contains | | |
| <ul style="list-style-type: none"> • 1 sample collection guide for pet owners • 3 compostable feces bag • 1 protective bag large | 10 pc. | 19.10 |
| IVD Swine Oral Fluid Kit each kit contains | | |
| <ul style="list-style-type: none"> • 1 user manual • 1 chewing rope made of cotton appr. 70 cm Ø 16 mm with fastening rope appr. 150 cm Ø 3 mm • 1 pair of disposable gloves • 1 large plastic bag for oral fluid collection • 1 sample tube • 2 small plastic bags • 1 adhesive label | 1 pc. | 9.60 |
| | from 12 pc. each | 9.00 |

HUMANS

SEROLOGICAL TESTS (Antibody Detection)

Material: serum or blood without anticoagulants

Erysipelothrix rhusiopathiae (Erysipelas) ELISA°

Material: serum or blood without anticoagulants

Leptospira MAT°

Selected antigens of representative strains of different pathogenic serovars and serogroups

Material: serum or blood without anticoagulants

Testing for individual strains or serovars is possible after consultation

HUMANS

PCR TESTS (Antigen detection)

Leptospira realtime PCR

incl. differentiation of pathogenic *Leptospira* (subclades P1 and P2)

Material: body fluids or tissue depending on clinical manifestation

SARS-CoV-2 (Severe Acute Respiratory Syndrome Coronavirus Type 2, COVID-19 pathogen) realtime PCR°

Material: nasal swabs, pharyngeal swabs, nasopharyngeal swabs, BALF, TBS, sputum, gargle sample

Pool examinations by epidemiological units are also possible Pool formation per sample (human pathogenic)

also in dogs, cats, rabbits, golden hamsters, ferrets

All examinations can be carried out as a service for self-paying patients or private patients.

For the collection of gargle samples, we offer practical sets including suitable packaging and shipping material.

Find more information on our website at <https://www.ivd-gmbh.de/home/>

Test methods offered for various pathogen detections in pigs

| Test methods / alphabetical list of pathogens | Indirect Pathogen Detection (Antibodies) | | | | | | Direct Pathogen Detection | | | | | | | |
|---|--|----------------|-----|----|-----|-----|---------------------------|------|-----------------|--------|------------|---------------|--------------|-----|
| | ELISA | Serotyp. ELISA | CFT | HI | MAT | RBT | PCR | qPCR | Differentiation | Typing | Sequencing | Bact. Culture | Parasitology | IHC |
| <i>Actinobacillus pleuropneumoniae</i> | • | • | | | | | • | | | • | | • | | • |
| ADV (Aujeszky's Disease Virus) | • | | | | | | | | | | | | | |
| ASFV (African Swine Fever Virus) | • | | | | | | | | | | | | | |
| <i>Bordetella bronchiseptica</i> | | | | | | | | | | | | • | | |
| <i>Brachyspira</i> species | | | | | | | • | | • | | | • | | |
| <i>Brachyspira hyodysenteriae</i> | | | | | | | • | | | | | • | | |
| <i>Brucella</i> , <i>Brucella abortus</i> | | | • | | | • | | | | | | | | |
| <i>Campylobacter</i> spec. | | | | | | | • | | | | | | | |
| Chlamydia | | | | | | | • | | | | | | | |
| Classical Swine Fever Virus (CSFV) | • | | | | | | | | | | | | | |
| <i>Clostridium perfringens</i> | | | | | | | • | | • | | | • | | |
| <i>Clostridioides difficile</i> | | | | | | | • | | • | | | • | | |
| Coronavirus (PEDV, TGEV) | | | | | | | • | | • | | | | | |
| <i>Cryptosporidia</i> | | | | | | | | | | | | | • | |
| <i>Cystoisospora suis</i> | | | | | | | • | | | | | | • | |
| <i>Enterococcus hirae</i> , <i>durans</i> or <i>villorum</i> | | | | | | | | | •* | | | • | | |
| <i>Escherichia coli</i> | | | | | | | • | | • | | | • | | |
| <i>Eperythrozoon (Mycoplasma) suis</i> | | | | | | | • | | | | | | | |
| <i>Erysipelothrix rhusiopathiae</i> (Erysipelas) | • | | | | | | | | | | | • | | |
| <i>Fusobacterium necrophorum</i> | | | | | | | | | | | | • | | |

| Test methods / alphabetical list of pathogens | Indirect Pathogen Detection (Antibodies) | | | | | | Direct Pathogen Detection | | | | | | | |
|--|--|----------------|-----|----|-----|-----|---------------------------|------|-----------------|--------|------------|---------------|--------------|-----|
| | ELISA | Serotyp. ELISA | CFT | HI | MAT | RBT | PCR | qPCR | Differentiation | Typing | Sequencing | Bact. Culture | Parasitology | IHC |
| <i>Glaesserella parasuis</i> | • | | | | | | • | | | •* | | • | | |
| Influenza A Virus (swIAV) | • | | | • | | | • | | | • | • | | | • |
| <i>Lawsonia intracellularis</i> (PIA) | • | | | | | | • | • | | | | | | • |
| Leptospira (pathogenic Serovars) | | | | | • | | • | | • | | | | | (•) |
| <i>Mesomycoplasma flocculare</i> | | | | | | | •* | | | | | • | | |
| <i>Mesomycoplasma hyopneumoniae</i> | • | | | | | | • | | | | | • | | • |
| <i>Mesomycoplasma hyorhinis</i> | • | | | | | | • | | | | | • | | |
| <i>Metamycoplasma hyosynoviae</i> | | | | | | | • | | | | | • | | |
| Parasite stages in feces, skin parasites (mites, lice) | | | | | | | | | | | | | • | |
| <i>Pasteurella multocida</i> toxin A | | | | | | | • | | | •* | | • | | |
| Porcine Circovirus 2 | • | | | | | | • | • | | | • | | | • |
| Porcine Circovirus 3 | | | | | | | • | | | | | | | |
| PEDV (Porcines Epidemic Diarrhoea Virus) | | | | | | | • | | | | | | | |
| Porcine Parvovirus | • | | | | | | • | | | | • | | | |
| PRRSV | • | | | | | | • | | • | | • | | | |
| Rotavirus Group A | | | | | | | • | | | | | | | • |
| Rotavirus Group C | | | | | | | • | | | | | | | |
| Salmonella | • | | | | | | • | | | •* | | • | | |
| <i>Schaalia (Actinomyces) hyovaginalis</i> | | | | | | | | | | | | • | | |
| <i>Sarcoptes scabiei</i> var. <i>suis</i> (Scabies Swine) | • | | | | | | | | | | | | • | |
| <i>Staphylococcus aureus</i> - MRSA | | | | | | | | | | •* | | • | | |
| <i>Staphylococcus chromogenes</i> | | | | | | | | | | •* | | • | | |
| <i>Staphylococcus hyicus</i> - Toxin detection | | | | | | | | | | •* | | • | | |
| <i>Streptococcus suis</i> | | | | | | | • | | | •* | | • | | |
| SVDV (Swine Vesicular Disease Virus) | (•) | | | | | | | | | | | | | |

| Test methods / alphabetical list of pathogens | Indirect Pathogen Detection (Antibodies) | | | | | | Direct Pathogen Detection | | | | | | | |
|--|--|----------------|-----|----|-----|-----|---------------------------|------|-----------------|--------|------------|---------------|--------------|-----|
| | ELISA | Serotyp. ELISA | CFT | HI | MAT | RBT | PCR | qPCR | Differentiation | Typing | Sequencing | Bact. Culture | Parasitology | IHC |
| TGEV (Transmissible Gastroenteritis Virus) | (•) | | | | | | • | | | | | | | |
| <i>Trueperella abortus</i> | | | | | | | | | | | | • | | |
| <i>Trueperella pyogenes</i> | | | | | | | | | | | | • | | |

* Examination only possible from isolates; (•) Examination on request

Suitable test materials for direct pathogen detection using PCR, bacteriology, histology and immunohistochemistry in pigs and ruminants

| suitable material / alphabetical list of pathogens | Meninges / Liquor | | Conjunctival swab | Blood / Milk ¹ | Nasal swabs | BALF/ TBS | Bronchial swabs | Lung | Tonsils/ Lymphnode | Feces / (fecal swabs) | Small intestine | Caecum / Colon | Cervical swabs | Abortion (Fetus/Placenta) | Serosa swabs | Joint | others |
|--|--|---|-------------------|---------------------------|-------------|-----------|-----------------|------|--------------------|-----------------------|-----------------|----------------|----------------|---------------------------|--------------|-------|---|
| | <i>Actinobacillus pleuropneumoniae</i> | | | | | | • | | • | • | | | | | | | |
| <i>Bordetella bronchiseptica</i> | | | | | • | • | • | • | | | | | | | | | |
| <i>Brachyspira species</i> | | | | | | | | | | • | | • | | | | | |
| <i>Brachyspira hyodysenteriae</i> | | | | | | | | | | • | | • | | | | | |
| <i>Campylobacter spec.</i> | | | | | | | | | | • | • | | | | | | |
| <i>Chlamydia</i> | | • | | | | | | | | | | | • | • | | | |
| <i>Clostridioides difficile</i> | | | | | | | | | | • | • | • | | | | | |
| <i>Clostridium perfringens</i> | | | | | | | | | | • | • | | | | | | |
| <i>Cystoisospora suis</i> | | | | | | | | | | • | | | | | | | swabs |
| Cryptosporidia | | | | | | | | | | • | • | • | | | | | |
| <i>Enterococcus durans</i> | | | | | | | | | | • | • | | | | | | |
| <i>Enterococcus hirae</i> | | | | | | | | | | • | • | | | | | | |
| <i>Enterococcus villorum</i> | | | | | | | | | | • | • | | | | | | |
| <i>Escherichia coli</i> | | | | | | | | | | • | • | • | | | | | Sepsis: kidney, CNS |
| <i>Eperythrozoon (Mycoplasma) suis</i> | | | | • | | | | | | | | | | | | | |
| <i>Erysipelothrix rhusiopathiae</i> (Erysipelas) | | | | | | | | | | | | | | | | • | kidney |
| <i>Fusobacterium necophorum</i> | | | | | | | | | | | | | | • | • | | abscess, claw swab |
| <i>Glaesserella parasuis</i> | | | | | | • | • | • | | | | | | | • | • | |
| Influenza A Virus | | | | | • | • | • | • | | | | | | | | | |
| <i>Lawsonia intracellularis</i> (PIA) | | | | | | | | | | • | • | • | | | | | |
| Leptospira (pathogenic Serovars) | • | | | • | | | | | | | | | • | • | | | kidney, liver, internal genital organs, body fluids, vitreous, aqueous humor, urine |
| <i>Mesomycoplasma hyopneumoniae</i> | | | | | | • | • | • | | | | | | | | | |

| suitable material / alphabetical list of pathogens | Meninges / Liquor | | Conjunctival swab | Blood / Milk ¹ | Nasal swabs | BALF/ TBS | Bronchial swabs | Lung | Tonsils/ Lymphnode | Feces / (fecal swabs) | Small intestine | Caecum / Colon | Cervical swabs | Abortion (Fetus/Placenta) | Serosa swabs | Joint | others |
|--|---------------------------------|--|-------------------|---------------------------|-------------|-----------|-----------------|------|--------------------|-----------------------|-----------------|----------------|----------------|---------------------------|--------------|-------|---------------|
| | <i>Mesomycoplasma hyorhinis</i> | | | | | • | • | • | | | | | | | | • | • |
| <i>Metamycoplasma hyosynoviae</i> | | | | | | | | | | | | | | | | • | |
| <i>Mesomycoplasma flocculare</i> | | | | | | | | • | | | | | | | | | |
| <i>Pasteurella multocida</i> | | | | | • | • | • | • | | | | | | | | | |
| Porcine Circovirus 2/3 | | | • | | • | | • | • | • | | • | • | • | • | | | |
| Porcine Coronavirus | | | | | | | | | | • | • | • | | | | | |
| Porcine Parvovirus | | | • | | | | | | | | | | | • | | | |
| Porcine Epidemic Virus | | | | | | | | | | • | • | | | | | | |
| PRRSV | | | • | • | • | • | • | • | | | | | | • | | | |
| Rotavirus Group A / C | | | | | | | | | | • | • | | | | | | |
| <i>Salmonella</i> | | | | | | | | | | • | • | • | | | | | |
| <i>Schaalia (Actinomyces) hyovaginalis</i> | | | | | | | | | | | | | • | • | | | abscess |
| <i>Staphylococcus aureus</i> - MRSA | • | | • ¹ | • | • | • | • | • | | | | | • | • | | • | abscess |
| <i>Staphylococcus chromogenes</i> | | | | | | | | | | | | | | | | | abscess, skin |
| <i>Staphylococcus hyicus</i> - toxin determination | | | | | | | | | | | | | • | • | | | abscess, skin |
| <i>Streptococcus dysgalactiae</i> | • | | • ¹ | • | • | • | • | • | | | | | • | • | | • | abscess |
| <i>Streptococcus suis</i> | • | | | | • | • | • | • | | | | | | | • | • | |
| Transmissible Gastroenteritis Virus | | | | | | | | | | • | • | | | | | | |
| <i>Trueperella abortusuis</i> | | | | | | | | | | | | | • | • | | | |
| <i>Trueperella pyogenes</i> | | | | | | | | • | | | | | • | • | | • | abscess |

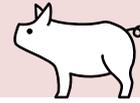
Multiplex PCR and combinations of different examination methods

| suitable material / pathogens | BALF/ TBS | Lung | Dry swab of Joints / Serosa | Feces |
|--|-----------|------|-----------------------------|-------|
| Multiplex PCRs | | | | |
| „Neumünster“ (<i>M. hyopneumoniae</i> , <i>M. hyorhinis</i> , swIAV, PCMV, PCV2, PRCV, PRRSV-1,-2) | ● | ● | | |
| „App / GPS + vir“ (<i>App</i> , <i>Gps</i>) | ● | ● | | |
| „Serositis“ (<i>Gps</i> , <i>M. hyorhinis</i>) | | | ● | |
| „Arthritis“ (<i>Gps</i> , <i>M. hyorhinis</i> , <i>M. hyosynoviae</i> , <i>Sc. suis</i>) | | | ● | |
| Combinations of different examination methods | | | | |
| „Outdoor Pigs - feces“ PCR + Parasitology (<i>Brachyspira</i> differentiation by PCR, <i>Laws. intracellularis</i> PCR, <i>Salmonella</i> Choleraesuis/ Typhimurium PCR; flotation) | | | | ● |
| „Saugferkeldurchfall/ suckling piglet diarrhoea“ Bact. culture + PCR +Parasitology (<i>E. coli</i> , <i>C. perfringens</i> , <i>Enterococcus</i> , Rota-/ Coronavirus; flotation) | | | | ● |

SWINE

SEROLOGICAL TESTS (Antibody detection)

Material: serum or blood without anticoagulants



ADV (Aujeszky's disease virus or pseudorabies virus (PRV),
Suid herpesvirus 1) ELISA

App (*Actinobacillus pleuropneumoniae*) ApxIV Toxin ELISA (standard)

App ApxII toxin ELISA (for research purpose only)

App LPS mix ELISA

App serotyping / single App serotypes / groups

- **App** ELISA virulent plus screening:
virulent serotypes (ST 1/9/11; 5a/b) plus ST 2 (dominant in EU)
- **App** ELISA full screening (12 serotypes)

- **App** ELISA serotypes 1/9/11
- **App** ELISA serotype 2
- **App** ELISA serotypes 3/6/8
- **App** ELISA serotypes 4/7
- **App** ELISA serotypes 5a/b
- **App** ELISA serotype 10
- **App** ELISA serotype 12

ASFV (African swine fever virus) ELISA
on request

Brucella spec. RBT

CSFV (Classical swine fever virus) ELISA

Erysipelothrix rhusiopathiae (Erysipelas) ELISA

Glaesserella parasuis (Gps) ELISA

Influenza A Virus ELISA (serum) (standard)

Influenza A Virus ELISA (oral fluid, saliva)

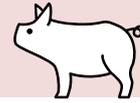
Influenza A Virus HI

(current strains of relevant subtypes in Germany;
recommended for vaccinated sows)

SWINE

SEROLOGICAL TESTS (Antibody detection)

Material: serum or blood without anticoagulants



Lawsonia intracellularis („PIA“) ELISA

Leptospira MAT

(Selected antigens of representative strains of different pathogenic serovars and serogroups)

Testing for individual strains or serovars is possible after consultation

Mesomycoplasma hyopneumoniae ELISA (standard)

Mesomycoplasma hyopneumoniae ELISA (alternative)

Mesomycoplasma hyorhinis ELISA

PCV2 (Porcine circovirus 2) IgM / IgG ELISA (standard)

PCV2 (Porcine circovirus 2) ELISA (alternative)

PPV (Porcine parvovirus) ELISA

PRRSV (Porcine reproductive and respiratory syndrome virus) ELISA (serum)

Salmonella ELISA

Export of results into the Qualiproof® database

(as part of the salmonella monitoring and reduction program for pig production by QS Qualität und Sicherheit GmbH, Bonn, these charges are collected and passed on to Qualitype AG, Dresden, Germany)

Sarcoptes scabiei var. suis (Scabies) ELISA

SVDV (Swine vesicular disease virus) ELISA

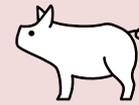
on request

TGEV (Transmissible gastroenteritis virus) /

PRCV (Porcine respiratory coronavirus) ELISA

on request

SWINE
EXAMINATION FOR INTERNATIONAL TRADE



Price on
request

Individually combined according to your specifications, e.g.:

African swine fever (ASFV)
Classical swine fever (CSFV, Hog Cholera)
Aujeszky's disease (ADV or PRV or SHV1)
Transmissible gastroenteritis (TGEV)
Swine vesicular disease (SVDV)
Brucellosis
PRRSV
and many more.

SWINE
SEROLOGICAL SCREENINGS
Material: Serum or blood without anticoagulants



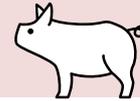
RESPIRATORY PATHOGENS

Respiratory tract screening "Ahlem"

Actinobacillus pleuropneumoniae
Mesomycoplasma hyopneumoniae
PRRSV
Influenza A Virus (ELISA) or
(HI)

Respiratory tract screening "Glässer"

Actinobacillus pleuropneumoniae
Glaesserella parasuis
PRRSV
Influenza A Virus (ELISA) or
(HI)



REPRODUCTIVE PATHOGENS

Reproductive tract screening “Ery + Parvo”

Erysipelothrix rhusiopathiae (Erysipelas)

Leptospira (pathogenic serovars)

PPV

PRRSV

Reproductive tract screening “Melle”

PCV2 IgM / IgG

PRRSV

Leptospira

Influenza A Virus (ELISA) or
(HI)

Screening „Sow Vaccination“

PCV2 (IgM/ IgG)

PRRSV

PPV

Influenza A Virus HI

Health Check “Swine”

Actinobacillus pleuropneumoniae

Erysipelothrix rhusiopathiae (Erysipelas)

Leptospira (pathogenic serovars)

Mesomycoplasma hyopneumoniae

PPV

PRRSV

Influenza A Virus (ELISA) or
(HI)

Screening „Outdoor Pigs“ - blood

Erysipelothrix rhusiopathiae (Erysipelas)

Leptospira (pathogenic serovars)

PRRSV

Influenza A Virus (ELISA)

SWINE PCR TESTS

directly from sample material



App (*Actinobacillus pleuropneumoniae*) apxIV PCR

Material: BALF, TBS, lung, tonsil

App capsule typing by multiplex PCR (cps mPCR) molecular „serotyping“ of *App* isolates

Material: only fresh BALF, TBS, lung, tonsil or fresh samples on FTA-Cards

Brachyspira hyodysenteriae realtime PCR

Material: feces, rectal swabs, large intestine

Brachyspira pilosicoli PCR

Material: feces, rectal swabs, large intestine

Brachyspira spec. PCR

Detection of *Brachyspira spec.*, *B. pilosicoli*, *B. hampsonii*, *B. intermedia*, *B. innocens*, *B. suanatina* and *B. murdochii*

Material: feces, rectal swabs, large intestine

Brachyspira differentiation PCR

Detection of *Brachyspira spec.*, *B. pilosicoli*, *B. hampsonii*, *B. intermedia*, *B. innocens*, *B. suanatina* and *B. murdochii* by PCR incl. *Brachyspira hyodysenteriae* by realtime PCR

Material: feces, rectal swabs, large intestine

Campylobacter differentiation PCR

Detection of *Campylobacter spec.*, *C. coli* and *C. jejuni*

Material: feces, rectal swabs, small intestine

Chlamydia PCR

Material: Genital swabs, conjunctival swabs, placenta, fetal liver, lung, spleen, genital organs

Clostridioides (Clostridium) difficile multiplex PCR

Detection of genes coding for toxin B and the binary toxin B

Material: feces, rectal swabs

Clostridium perfringens multiplex PCR

Detection of types A and C

Material: feces, rectal swabs

Cystoisospora suis PCR°

Material: feces, rectal swabs, environmental samples

Escherichia coli „Virulence-Associated Factors“ multiplex PCR

Detection of virulence associated factor genes (fimbriae, adhesins, toxins and other factors)

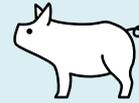
Material: feces, rectal swabs

Escherichia coli „EDEC“ multiplex PCR

Detection of EDEC- associated genes (shigatoxine 2e, F18-fimbriae),
Identification of the pathotype of edema disease

Material: feces, rectal swabs

SWINE
PCR TESTS
directly from sample material



Glaesserella parasuis (gps) multiplex PCR
Detection incl. potential marker genes of virulence
Material: BALF, TBS, lung

Influenza A virus (swIAV) realtime PCR
Material: Nasal swabs (BALF, TBS, oral fluid/saliva), lung

Influenza A virus (swIAV) realtime multiplex PCR for subtyping of
European swine influenza A virus
Material: Nasal swabs (BALF, TBS, oral fluid/saliva), lung

Influenza A virus (swIAV) Sequencing° (by external lab) of a PCR product
(HA gene of influenza A virus) incl. comparison and alignment to a reference
strain
PCR
Sequencing and alignment
total

Allocation of sequence files

Lawsonia intracellularis realtime PCR
Material: feces, rectal swabs, small intestine

Lawsonia intracellularis quantitative PCR (only individual samples)
Material: feces, rectal swabs, small intestine

Leptospira realtime PCR
Detection including the differentiation of pathogenic *Leptospira*
(subclades P1 and P2)
Material: Genital swabs, urine, sperm, kidney, fetal tissues (kidney, liver, lung),
placenta, genital organs (for *L. Bratislava*)

Mesomycoplasma (Mycoplasma) hyopneumoniae realtime PCR
Material: BALF, TBS, nasal swabs, bronchus swabs, lung

Mesomycoplasma (Mycoplasma) hyorhinis PCR
Material: BALF, TBS, nasal swabs, lung, bronchus swab

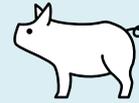
Metamycoplasma (Mycoplasma) hyosynoviae PCR
Material: synovial fluid, joint capsule, joint swab

Eperythrozoon (Mycoplasma) suis („eperythrozoonosis“) PCR
Material: blood (up to 5 samples may be pooled), (spleen, lung)

***Pasteurella multocida* Toxin (PMT)** PCR
Detection of *toxA*-gen
Material: nasal swabs, (up to 5 samples may be pooled), bacterial isolates

SWINE
PCR TESTS

directly from sample material



PCV2/3 (Porcine circovirus 2 and 3) realtime PCR

Material: BALF, TBS, blood, oral fluid/ saliva, tonsil, lymph node
(up to 5 samples may be pooled)

PCV2 quantitative PCR (**only individual samples!**)

Material: BALF, TBS, blood, oral fluid/ saliva, tonsil, lymph node

PCV2 Sequencing° (by external lab) of PCR product (orf2)

incl. comparison of identity and alignment to the reference strain of the respective genotypes (type PCV2 a-h)

PCR

Sequencing and alignment

total

Allocation of sequence files

PPV (Porcine parvovirus) PCR

Material: abortion material, fetal lung

PPV Sequencing° (by external lab) of a PPV1 PCR product (VP2)

incl. comparison of identity and alignment to a reference strain

PCR

Sequencing and alignment

total

Allocation of sequence files (per data set)

PRRSV realtime multiplex PCR

differentiation of European and North American genotype or rather species
PRRSV-1 and PRRSV-2

Material: BALF, TBS, blood, oral fluid/ saliva, sperm (intermittent excretion), lung,
tonsil, placenta (up to 5 blood samples may be pooled)

PRRSV Sequencing° (by external lab) of PCR product

incl. comparison of identity and alignment to the reference strain of the
respective species (type 1 (EU): strain Lelystad or type 2 (NA): VR 2332 =
Ingelvac PRRS MLV (BI)) and vaccine strains registered in Germany

PCR

Sequencing and alignment

total

Alignment to further strains / sequences (per strain/ sequence)

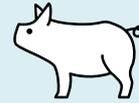
Allocation of sequence files (per strain/sequence)

Rotavirus group A and C realtime PCR

Material: feces, rectal swabs, small intestine

SWINE
PCR TESTS

directly from sample material



Salmonella Choleraesuis Typhimurium PCR

detection of *Salmonella* and differentiation of serovars Choleraesuis and Typhimurium after cultural enrichment

Material: feces, rectal swabs, large intestine; isolates from bacterial culture

Porcine corona virus PCR:

TGEV (Transmissible gastroenteritis virus) & **PEDV** (Porcine epidemic diarrhoea virus) PCR

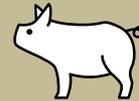
Material: feces, rectal swabs, small intestine

Streptococcus suis (Sc. suis) PCR

Material: synovial fluid, joint swab, joint capsule, meningeal swab, serosa

SWINE

**Combinations of different examination methods/
Diagnostic Profiles**



Combinations of different examination methods

Screening „Outdoor Pigs“ - feces

Material: only with feces samples, no swabs

Brachyspira differentiation PCR

Lawsonia intracellularis realtime PCR

Salmonella Choleraesuis Typhimurium PCR

Detection of parasites (flotation)

Diagnostic profil „Suckling piglet diarrhoea“

Material: only with feces samples, no swabs

Rota-/Coronavirus PCR

Bacterial culture for *E. coli*, *Enterococcus* and *C. perfringens*

Detection of parasites (flotation)

with add. bacterial culture for *C. difficile*

Optional, if positive:

Typing *E. coli* (multiplex PCR for detection of 21 virulence associated factors)

Typing *C. perfringens* (multiplex PCR & Immunoblot)

Identifying *Enterococcus* (PCR)

Resistance testing (Agar diffusion))

Resistance testing (Microdilution)

Resistance testing (Microdilution anaerobes)



RESPIRATORY DISEASES

PCR Screening „Respiration“ (Material: BALF, lung)

Actinobacillus pleuropneumoniae

Mesomycoplasma hyopneumoniae

Influenza A Virus

PRRSV (realtime PCR with differentiation PRRSV 1 (EU) and 2 (NA))

PCR Screening „Respiration plus “ (Material: BALF, lung)

Actinobacillus pleuropneumoniae

Mesomycoplasma hyopneumoniae

Influenza A Virus

PRRSV (realtime PCR with differentiation PRRSV 1 (EU) and 2 (NA))

PCV2/3

PCR Screening „Bakum“ (Material: BALF, lung)

Mesomycoplasma hyopneumoniae

Influenza A Virus

PCV2/3

PRRSV (realtime PCR with differentiation PRRSV 1 (EU) and 2 (NA))

PCR Screening „Dessau“ (Material: BALF, lung)

Actinobacillus pleuropneumoniae

Mesomycoplasma hyopneumoniae

PRRSV (realtime PCR with differentiation PRRSV 1 (EU) and 2 (NA))

PCV2/3

REPRODUCTIVE DISEASES

PCR Screening „Reproduction“

(Material: Pool of fetal lung, thymus, heart, kidney, placenta/ liver, genital swab)

Chlamydia

Leptospira (pathogenic)

PCV2/3

PPV

PRRSV (realtime PCR with differentiation PRRSV 1 (EU) and 2 (NA))

DIARRHOEA

PCR Screening „Dysentery / Ileitis“

(Material: feces, small and large intestine)

Brachyspira hyodysenteriae

Lawsonia intracellularis



PCR Screening „Enteritis“ (Material: feces, small and large intestine)

Brachyspira hyodysenteriae

Lawsonia intracellularis

Salmonella serovars Choleraesuis and Typhimurium

PCR Screening „Rota- / Coronavirus“ (Material: feces, small intestine)

Rotavirus group A and C

TGEV (Transmissible Gastroenteritis Virus)

PEDV (Porcine Epidemic Diarrhoea Virus)

PCR Screening* „Suckling piglet diarrhoea“ (Material: feces, small intestine)

E.coli incl.typing “virulence associated factors“

Clostridium perfringens incl. detection of types A and C

Cystoisospora suis^o

Rotavirus A/C

PCR Screening* „Suckling piglet diarrhoea plus“ (Material: feces, small intestine)

E.coli incl.typing “virulence associated factors“

Clostridium perfringens incl. detection of types A and C

Cystoisospora suis^o

Rotavirus A/C

TGEV/PEDV)

PCR Screening* „Finisher“ (Material: feces, small and large intestine)

E.coli incl.typing “virulence associated factors“

Brachyspira hyodysenteriae

Lawsonia intracellularis

Salmonella serovars Choleraesuis and Typhimurium

PCR Screening* „Finisher plus“ (Material: feces, small and large intestine)

E.coli incl.typing “virulence associated factors“

Brachyspira hyodysenteriae

Lawsonia intracellularis

Salmonella serovars Choleraesuis and Typhimurium

Rotavirus A/C

TGEV/PEDV

***Please note:**

In the PCR screenings “Suckling piglet diarrhoea” and “Finisher” there is no pathogen cultivation, a resistance test and the preservation of isolates are not possible without further cultural examination!



PCR Screening „Oral Fluids“ for Herd Check

(Material: oral fluid)

Mesomycoplasma (Mycoplasma) hyopneumoniae

Mesomycoplasma (Mycoplasma) hyorhinis

Influenza A Virus

PCMV

PCV2

PRCV

PRRSV 1 and 2 (EU/NA)

PPV

PEDV

Rotavirus A/C

Brachyspira hyodysenteriae

Lawsonia intracellularis

MULTIPLEX PCR - SWINE

Multiplex PCR „Neumünster“ (Material: BALF, TBS, lung)

Mesomycoplasma hyopneumoniae (incl. confirming PCR)

Mesomycoplasma hyorhinis

Influenza A Virus

PCMV

PCV2

PRCV

PRRSV 1 and 2 (EU/NA)

Multiplex PCR „APP / GPS (HPS) + vir“ (Material: BALF, TBS, lung)

Actinobacillus pleuropneumoniae

Glaesserella parasuis + potential virulence factor

Multiplex PCR „Serositis“ (Material: dry swabs from serosa)

Glaesserella parasuis + potential virulence factor

Mesomycoplasma hyorhinis

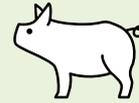
Multiplex-PCR "Arthritis" (Material: joint swabs, joint capsule)

Glaesserella parasuis

Mesomycoplasma hyorhinis

Metamycoplasma hyosynoviae

Streptococcus suis



Examination by bacteriological culture, basic

Examination by bacteriological culture for respiratory pathogens
(incl. necessary selective culture media)

Examination by bacteriological culture incl. anaerobes
using enrichment and selective culture media

Examination by bacteriological culture for *Brachyspira*
using selective culture media

Examination by culture only for *Salmonella* according
DIN EN ISO 6579 1 using enrichment and selective culture media

Examination by bacteriological culture incl. *Salmonella* according
DIN EN ISO 6579 1 using enrichment and selective culture media

Examination by culture for *E. coli*, *C. perfringens* and *Salmonella*
according DIN EN ISO 6579 1:2017 using enrichment and selective culture
media

Examination by bacteriological culture incl. *Mycoplasma*
using enrichment and selective culture media
Note: special transport medium is available on request

Additional selective culture medium

Please note that the identification of pathogenic bacteria species may incur further costs for diagnostic verification, e.g. for species identification by PCR or 16S rRNA typing (PCR and sequencing° (by external lab) including sequence analysis).

microbiological disinfection check°
Total bacterial count and viable count *E. coli*, *C. perfringens* and mold fungi
Note: pre-register one week in advance, please

microbiological disinfection check°
only total bacterial count
Note: pre-register one week in advance, please

Most suitable sample materials for cultural detection of bacterial pathogens of:

| | |
|-------------------------------|--|
| Arthritis | swab from joint capsule or joint cartilage, joint capsule, (synovia) |
| Respiratory infections | lung, lung and bronchial swab, bronchoalveolar lavage fluid (BALF) |
| Dermatitis | skin, deep skin scrapings |
| Endometritis | cervical swab |
| Enteritis, Diarrhoea | feces, fecal swab, rectal swab, small and large intestine |
| Meningitis | meningeal swab, CSF (liquor) |
| Rhinitis | nasal swab |
| Sepsis | kidney, spleen |
| Serositis | serosal swab |

Note: **swabs with medium** are most suitable for cultural examination

SWINE BACTERIOLOGY



Identification of bacteria isolated at IVD as part of the cultural examination

Species identification by PCR of:

Actinobacillus pleuropneumoniae

Bordetella bronchiseptica

Clostridioides difficile

Enterococcus durans

Enterococcus hirae

Erysipelothrix rhusiopathiae (Erysipelas)

Fusobacterium necrophorum

Glaesserella parasuis

Mesomycoplasma flocculare

Mesomycoplasma hyopneumoniae

Mesomycoplasma hyorhinis

Metamycoplasma hyosynoviae

Pasteurella multocida

Schaalia (Actinomyces) hyovaginalis

Staphylococcus chromogenes

Staphylococcus hyicus

Streptococcus dysgalactiae

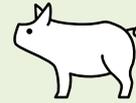
Streptococcus suis

Trueperella abortus

16S rRNA sequencing° (for identification of any bacterial species)

ITS sequencing° (for identification of further mycological species)

Cultural biochemical or serological differentiation



Resistance / Susceptibility testing of bacterial isolates

Agar diffusion test

Microdilution technique

(determination of the minimal inhibitory concentrations (MICs))

Microdilution technique anaerobes

Custodial storage of bacterial isolates/ Mycoplasma

Delivery of characterised bacterial field isolates

e.g. *Mycoplasma*, *Glaesserella parasuis* et al.

Identification / differentiation of submitted bacterial isolates

by PCR

Actinobacillus pleuropneumoniae

Bordetella bronchiseptica

Clostridioides difficile

Erysipelothrix rhusiopathiae (Erysipelas)

Fusobacterium necrophorum

Glaesserella parasuis

Mesomycoplasma flocculare

Mesomycoplasma hyopneumoniae

Mesomycoplasma hyorhinis

Metamycoplasma hyosynoviae

Pasteurella multocida

Schaalia (Actinomyces) hyovaginalis

Staphylococcus chromogenes

Streptococcus dysgalactiae

Streptococcus suis

Trueperella abortusis

Additional subculture for submitted bacterial isolates

for further analysis

16S rRNA sequencing° (for identification of any bacterial species)

ITS sequencing° (for identification of further mycological species)



Typing of bacterial isolates/

Detection & typing of preserved nucleid acid from suitable samples

***Actinobacillus pleuropneumoniae* PCR**

Apx toxin typing

***Actinobacillus pleuropneumoniae* multiplex PCR**

Determination of the serotype

***Clostridioides (Clostridium) difficile* PCR**

Detection of the genes coding for toxin A and B and the binary toxins A and B

Clostridium perfringens

Determination of types A to F with/without β 2 toxin gene/enterotoxin gene by PCR and detection of α and β 2 toxin production by immunoblot

only: PCR

***Enterococcus (E.) hirae, E. durans, E. villorum* multiplex PCR**

Detection and differentiation of *E. hirae*, *E. durans* and/ or *E. villorum*

***E. coli* “Virulence-Associated Factors” multiplex PCR**

Detection of virulence-associated factor genes (fimbriae, adhesins, toxins, and other factors) incl. Edema Disease

***Glaesserella parasuis* / Gps Typing by PCR**

Serotyping by multiplex PCR Assay (serotypes 1-15) and Pathotyping (vtaA-LS-PCR)

only: Determination of the **Gps serotype** by multiplex PCR Assay (serotypes 1-15)

only: **Pathotyping** of **Gps** (prognosis of the possible virulence potential) by vtaA-LS-PCR

***Pasteurella multocida* toxin PCR**

Detection of the *tox*A gene

***Pasteurella multocida* capsule typing multiplex PCR**

Detection of capsular type A, B, D, E and F as well as haemorrhagic septicaemic (HS) capsular type B

***Salmonella* Serotyping** (according to the Kauffmann-White scheme)

Classification into serogroups A-E and F-67 by agglutination

***Salmonella Choleraesuis* Typhimurium multiplex PCR**

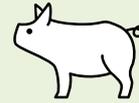
Detection of *Salmonella* and differentiation of serovar Choleraesuis and Typhimurium

Differentiation of *Salmonella* spp. vaccine and field strains

Differentiation of the vaccine strains from Salmoporc

***Staphylococcus aureus*, methicillin resistant s. MRSA**

SWINE BACTERIOLOGY



Staphylococcus hyicus multiplex PCR
Detection of exfoliative toxin genes A-D and shetaA

Streptococcus suis multiplex PCR
Determination of capsular type (cps 1, 2, 4, 7 or 9) and virulence-associated factor genes

Detection of resistance markers in bacterial isolates

mcr-1 gene (colistin resistance) PCR
Detection of mcr-1 gene, which confers resistance to colistin

MRSA Methicillin resistant *Staphylococcus aureus*
Detection of mecA gene by PCR and of expression of penicillin-binding protein 2 (PBP2) by agglutination

SWINE PARASITOLOGY°



Detection of parasite stages in feces Sedimentation Flotation
Helminth eggs, protozoan oocysts and cysts
Material: at least 10 g feces

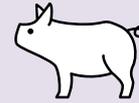
Cryptosporidia Staining according to Heine
Semi-quantitative direct detection of *Cryptosporidium* oocysts
Material: 2-5 g feces (if possible, fresh and cooled, but not frozen!) or fecal smear

***Cystoisospora suis* PCR°**
Material: feces, environmental samples

Detection of skin parasites:

Mites Microscopy, KOH procedure
Sarcoptes (Scabies), Demodex (Demodicosis)
Material: deep skin scrapings

Lice (*Haematopinus suis*)
Microscopy
Material: 3 transparent adhesive film strips per animal on microscope slides, combs and skin material



NECROPSY of animals up to 59 kg bodyweight including disposal costs

Animal carcasses are disposed of in accordance with the Animal Carcass Disposal Act according to the tariffs applicable in Lower Saxony, Germany.

Fetus and piglet (up to 5 kg) incl. placenta

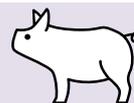
Young pig (5 - 15 kg)

Young pig (16 - 29 kg)

Pig (30 - 59 kg)

Removal of specimen from aborted fetuses

per sample



Histological examination (standard staining)

with report

Special stainings (Ziehl-Neelsen, Gram, etc.)

IMMUNOHISTOLOGY

***Actinobacillus pleuropneumoniae* IHC**

Material: Lung, formalin fixed

Influenza A Virus IHC

Material: Lung (cranioventral parts mit bronchus), formalin fixed

***Lawsonia intracellularis* IHC**

Material: Ileum, formalin fixed

***Mesomycoplasma (Mycoplasma) hyopneumoniae* IHC**

Material: Lung (cranioventral parts mit bronchus), formalin fixed

PCV2 IHC

Material: Lymph node, lung, kidney, spleen, GI-tract, formalin fixed

Rotavirus group A IHC

Material: small intestine, formalin fixed

Information for sampling technique

Tissue samples for histological and immunohistological examinations should be sent in at least **10 times the volume** of 4 or 10% fixed formalin and should not be thicker than **1 cm** in at least one dimension to ensure a sufficiently rapid penetration with the fixing solution.

A cover letter with a clinical preliminary report is essential for the interpretation of the histological findings and the limitation of the deducible etiological differential diagnoses.

Immunohistological examinations for the specific detection of certain pathogens in the tissue are only useful, if a previous histological examination revealed morphological changes, which justify the suspicion of infection with the pathogen in question. Is the meaning, which role a pathogen has played in a specific disease process, not of interest but just the question, whether or not an animal was infected with the pathogen, a PCR analysis of unfixed tissue is preferable due to its higher sensitivity.

Test methods offered for various pathogen detections in ruminants

| Test methods / alphabetical list of pathogens | Indirect Pathogen Detection (Antibodies) | | | | | | Direct Pathogen Detection | | | | | | | |
|---|--|----------------|-----|----|-----|-----|---------------------------|------|-----------------|--------|------------|---------------|--------------|-----|
| | ELISA | Serotyp. ELISA | CFT | HI | MAT | RBT | PCR | qPCR | Differentiation | Typing | Sequencing | Bact. Culture | Parasitology | IHC |
| Bovine Coronavirus | | | | | | | • | | | | | | | |
| <i>Brucella</i> , <i>Brucella abortus</i> | | | • | | | • | | | | | | | | |
| <i>Campylobacter spec.</i> | | | | | | | • | | | | | | | |
| Chlamydia | | | | | | | • | | | | | | | |
| <i>Clostridium perfringens</i> | | | | | | | | | •* | | • | | | |
| <i>Corynebacterium pseudotuberculosis</i> | • | | | | | | | | | | • | | | |
| <i>Coxiella burnetii</i> | (•) | | | | | | | | | | | | | |
| <i>Cryptosporidia</i> | | | | | | | | | | | | • | | |
| <i>Escherichia coli</i> | | | | | | | | | •* | | • | | | |
| <i>Erysipelothrix rhusiopathiae</i> (Erysipelas) | | | | | | | | | | | • | | | |
| <i>Fusobacterium necrophorum</i> | | | | | | | | | | | • | | | |
| <i>Histophilus somni</i> | | | | | | | • | | | | • | | | |
| Fasciola, Dicrocoelium (Ruminants) | | | | | | | | | | | | • | | |
| Leptospira (pathogenic Serovars) | | | | | • | | • | • | | | | | | |
| <i>Mannheimia haemolytica</i> | | | | | | | • | | | | • | | | |
| <i>Mycobacterium avium ssp. paratuberculosis</i> | (•) | | | | | | • | | | | | | | |
| <i>Mycoplasma bovis</i> | (•) | | | | | | • | | | | • | | | |
| <i>Mycoplasma bovirhinis</i> | | | | | | | | | | | • | | | |
| <i>Mesomycoplasma ovipneumoniae</i> | | | | | | | | | | | • | | | |
| <i>Neospora caninum</i> | • | | | | | | | | | | | | | |
| Parasite stages in feces, skin parasites (mites, lice) | | | | | | | | | | | | • | | |
| <i>Pasteurella multocida</i> | | | | | | | | | •* | | • | | | |
| Rotavirus Group A | | | | | | | • | | | | | | | • |

| Test methods / alphabetical list of pathogens | Indirect Pathogen Detection (Antibodies) | | | | | | Direct Pathogen Detection | | | | | | | |
|--|--|----------------|-----|----|-----|-----|---------------------------|------|-----------------|--------|------------|---------------|--------------|-----|
| | ELISA | Serotyp. ELISA | CFT | HI | MAT | RBT | PCR | qPCR | Differentiation | Typing | Sequencing | Bact. Culture | Parasitology | IHC |
| Rotavirus Group C | | | | | | | • | | | | | | | |
| <i>Staphylococcus aureus</i> - MRSA | | | | | | | | | | •* | | • | | |
| <i>Staphylococcus chromogenes</i> | | | | | | | | | | | | • | | |
| <i>Streptococcus dysgalactiae</i> | | | | | | | | | | | | • | | |
| <i>Streptococcus parauberis</i> | | | | | | | | | | | | • | | |
| <i>Streptococcus uberis</i> | | | | | | | | | | | | • | | |
| <i>Trueperella pyogenes</i> | | | | | | | | | | | | • | | |

* Examination only possible from isolates () examination on request

Suitable test materials for direct pathogen detection using PCR, bacteriology, histology and immunohistochemistry in ruminants

| suitable material / alphabetical list of pathogens | Meninges / Liquor | | Conjunctival swab | Blood / Milk ¹ | Nasal swabs | BALF/ TBS | Bronchial swabs | Lung | Tonsils/ Lymphnode | Feces / (fecal swabs) | Small intestine | Caecum / Colon | Cervical swabs | Abortion (Fetus/Placenta) | Joint | Others |
|--|----------------------------|--|-------------------|---------------------------|-------------|-----------|-----------------|------|--------------------|-----------------------|-----------------|----------------|----------------|---------------------------|-------|---------|
| | <i>Campylobacter spec.</i> | | | | | | | | | | • | • | | | | |
| <i>Chlamydia</i> | | | • | | | | | | | | | | • | • | | |
| <i>Clostridiodes difficile</i> | | | | | | | | • | | | • | | • | | | |
| <i>Clostridium perfringens</i> | | | | | | | | | | • | • | • | | | | |
| bovine Corona Virus | | | | | • | • | • | • | | • | • | • | | | | |
| Cryptosporidia | | | | | | | | | | • | • | • | | | | |
| Endoparasites | | | | | | | | | | • | • | • | | | | |
| <i>Escherichia coli</i> | | | | | | | | | | • | • | • | | | | |
| <i>Erysipelothrix rhusiopathiae</i> (Erysipelas) | | | | | | | | | | | | | | | • | Kidney |
| <i>Fusobacterium necrophorum</i> | | | | | | | | | | | | | • | | | Abscess |
| <i>Histophilus somni</i> | • | | | | • | • | • | • | | | | | | | | |
| <i>Fasciola, Dicrocoelium</i> | | | | | | | | | | • | • | • | | | | |
| Leptospira (pathogenic Serovars) | | | | | | | | | | | | | • | • | | |
| <i>Mannheimia haemolytica</i> | | | | | • | • | • | • | | | | | | | | |
| <i>Mycoplasma bovis</i> | | | | • ¹ | • | • | • | • | | | | | | | • | |
| <i>Mycoplasma bovirhinis</i> | | | | | • | • | • | • | | | | | | | | |
| <i>Mesomycoplasma ovipneumoniae</i> | | | | | • | • | • | • | | | | | | | | |
| <i>Pasteurella multocida</i> | | | | | • | • | • | • | | | | | | | | |
| Rotavirus Group A / C | | | | | | | | | | • | • | | | | | |
| <i>Salmonella</i> | | | | | | | | | | • | • | • | | | | |
| <i>Staphylococcus aureus</i> (MRSA) | • | | | • ¹ | • | • | • | • | | | | | • | • | • | |
| <i>Staphylococcus chromogenes</i> | | | | • ¹ | | | | | | | | | | | | |
| <i>Streptococcus dysgalactiae</i> | | | | • ¹ | | | | | | | | | • | • | | |
| <i>Streptococcus parauberis</i> | | | | • ¹ | | | | | | | | | | | | |

| suitable material / alphabetical list of pathogens | Meninges / Liquor | Conjunctival swab | Blood / Milk ¹ | Nasal swabs | BALF/ TBS | Bronchial swabs | Lung | Tonsils/ Lymphnode | Feces / (fecal swabs) | Small intestine | Caecum / Colon | Cervical swabs | Abortion (Fetus/Placenta) | Joint | Others |
|--|-----------------------------|-------------------|---------------------------|----------------|-----------|-----------------|------|--------------------|-----------------------|-----------------|----------------|----------------|---------------------------|-------|---------|
| | <i>Streptococcus uberis</i> | | | ● ¹ | | | | | | | | | ● | | |
| <i>Trueperella pyogenes</i> | | | ● ¹ | ● | ● | ● | ● | | | | | ● | | ● | Abscess |

Multiplex PCRs Ruminants / Combinations of different examination methods

Cattle

| suitable material / pathogen | Feces | Small intestine |
|---------------------------------|---|-----------------|
| | “Diarrhoea of Calves” Bact. culture + PCR + Parasitology (<i>E. coli</i> , <i>C. perfringens</i> , Rota-/ Coronavirus; flotation, Cryptosporidia special staining) | ● |

| suitable material / pathogen | Nasal swab | BALF/ TBS | Bronchial swabs | Lung |
|---------------------------------|--|-----------|-----------------|------|
| | “Ruminant Respiratory Pathogens” (<i>M. bovis</i> , <i>H. somni</i> , <i>P. multocida</i> , <i>M. haemolytica</i> , bovine Coronavirus, BRSV, PI3) | ● | ● | ● |

SHEEP AND GOAT
SEROLOGICAL TESTS (Antibody detection)
Material: serum or blood without anticoagulants



Brucella spec. RBT

Corynebacterium pseudotuberculosis ELISA

Coxiella burnetii (Q-Fever) ELISA

Material: blood serum, milk (individual and tank milk samples)
on request

Erysipelothrix rhusiopathiae (Erysipelas) SLA

Leptospira MAT

(Selected antigens of representative strains of different pathogenic serovars
and serogroups)

Testing for individual strains or serovars is possible after consultation

SHEEP AND GOAT
PCR TESTs (Antigen detection)



Leptospira realtime PCR

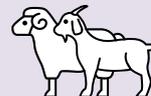
Detection including the differentiation of pathogenic *Leptospira*
(subclades P1 and P2)

Material: Genital swabs, urine, sperm, kidney, fetal tissues (kidney, liver, lung),
placenta, genital organs (for *L. Bratislava*)

Mycobacterium avium* subsp. *paratuberculosis PCR

Material: feces, fecal swabs, milk, mesenterial lymph node, small intestine

SHEEP AND GOAT
HISTOLOGY UND IMMUNHISTOLOGY



Histological examination (standard staining)

Special stainings (Ziehl-Neelsen, Gram, etc.)

**Necropsy of small ruminants is not possible due to animal health
legislation regulations.**



Examination by bacteriological culture, basic

Examination by bacteriological culture incl. anaerobes
using enrichment and selective culture media

Examination by bacteriological culture only *Salmonella* according
DIN EN ISO 6579 1 using enrichment and selective culture media

Examination by bacteriological culture incl. *Salmonella* according
DIN EN ISO 6579 1 using enrichment and selective culture media

**Examination by bacteriological culture: *E.coli*, *Cl. perfringens* and
*Salmonella*** according DIN EN ISO 6579 1 using enrichment and selective
culture media

Examination by bacteriological culture incl. *Mycoplasma*
using enrichment and selective culture media
Note: special transport medium is available on request

Additional selective culture medium

Please note that the identification of pathogenic bacteria species may incur further costs for diagnostic verification, e.g. for species identification by PCR or 16S rRNA typing (PCR and sequencing° (by external lab) including sequence analysis).

Most suitable sample materials for cultural detection of bacterial pathogens of:

| | |
|-------------------------------|--|
| Arthritis | swab from joint capsule or joint cartilage, joint capsule, (synovia) |
| Respiratory infections | lung, lung and bronchial swab, bronchoalveolar lavage fluid (BALF) |
| Dermatitis | skin, deep skin scrapings |
| Endometritis | cervical swab |
| Enteritis, Diarrhoea | feces, fecal swab, rectal swab, small and large intestine |
| Meningitis | meningeal swab, CSF (liquor) |
| Rhinitis | nasal swab |
| Sepsis | kidney, spleen |
| Serositis | serosal swab |

Note: **swabs with medium** are most suitable for cultural examination



Identification of bacteria isolated at IVD as part of the cultural examination

Identification of species by PCR:

Clostridioides difficile

Fusobacterium necrophorum

Histophilus somni

Mannheimia haemolytica

Mycoplasma mycoides (*Mycoplasma*) *bovirhinis*

Mesomycoplasma (*Mycoplasma*) *ovipneumoniae*

Pasteurella multocida

Streptococcus parauberis

Streptococcus uberis

16S rRNA sequencing° (for identification of any bacterial species)

ITS sequencing° (for identification of further mycological species)

Cultural biochemical or serological differentiation

Resistance / Susceptibility testing of bacterial isolates

Agar diffusion test

Microdilution technique

(determination of the minimal inhibitory concentrations (MICs))

Microdilution technique anaerobes

Custodial storage of bacterial isolates / mycoplasma

Delivery of characterised bacterial field isolates

e. g. *Corynebacterium pseudotuberculosis*, *Mycoplasma*, *Mannheimia*, *Pasteurella* and others

microbiological disinfection check°

Total bacterial count and viable count *E. coli*, *C. perfringens* and mold fungi

Note: pre-register one week in advance, please

microbiological disinfection check°

only total bacterial count

Note: pre-register one week in advance, please



Identification / differentiation of submitted bacterial isolates
by PCR

Clostridioides difficile
Fusobacterium necrophorum
Histophilus somni
Mannheimia haemolytica
Mycoplasma (*Mycoplasma*) *bovirhinis*
Mesomycoplasma (*Mycoplasma*) *ovipneumoniae*
Pasteurella multocida
Streptococcus parauberis
Streptococcus uberis

Additional subculture for submitted bacterial isolates
for further analysis

16S rRNA sequencing° (for identification of any bacterial species)

ITS sequencing° (for identification of further mycological species)

Typing of bacterial isolates

Clostridium perfringens

Determination of types A to F with/without β 2 toxin gene/enterotoxin gene
by PCR and detection of α and β 2 toxin production by immunoblot
only by PCR

***Escherichia coli* “Virulence associated Factors” PCR**

Detection of virulence-associated factor genes (fimbriae, adhesins,
toxins, and other factors)

MRSA Methicillin resistant *Staphylococcus aureus*

Detection of *mecA* gene by PCR and of expression of penicillin-binding
protein 2 (PBP2) by agglutination

***Pasteurella multocida* capsule typing PCR**

Detection of capsular type A, B, D, E and F as well as haemorrhagic
septicaemic (HS) capsular type B

***Salmonella* Serotyping** (according to the Kauffmann-White scheme)

Classification into serogroups A-E and F-67 by agglutination

***Staphylococcus aureus*, methicillin resistant s. MRSA**



Detection of parasite stages in feces Sedimentation and flotation

Helminth eggs, protozoan oocysts and cysts

Material: at least 10 g feces

Nematode larvae Baermann-Wetzel migration technique
of lung worms (*Dictyocaulus filaria*) and Protostrongylidae

Material: at least 10 g feces

Cryptosporidia Staining according to Heine

Semi-quantitative direct detection of *Cryptosporidium* oocysts

Material: 2-5 g feces (if possible, fresh and cooled, but not frozen!) or fecal smear

Liver and rumen fluke eggs (*Fasciola*, *Dicrocoelium*) sedimentation

Detection of eggs

Material: at least 10 g feces

Detection of skin parasites:

Mites Microscopy, KOH procedure

Sarcoptes, *Chorioptes*, *Psoroptes* mange, etc.

Material: deep skin scrapings

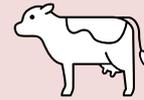
***Mallophaga*, lice** Microscopy

Material: 3 transparent adhesive film strips per animal on microscope slides, combings and skin material

CATTLE

SEROLOGICAL TESTS (Antibody detection)

Material: serum or blood without anticoagulants



Brucella spec. RBT

Coxiella burnetii (Q fever) ELISA

on request

Erysipelothrix rhusiopathiae (Erysipelas) SLA

Leptospira MAT

Selected antigens of representative strains of different pathogenic serovars and serogroups

Testing for individual strains or serovars is possible after consultation

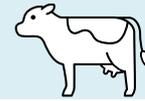
Mycobacterium avium* subsp. *paratuberculosis ELISA

on request

Mycoplasma* (*Mycoplasma*) *bovis ELISA

on request

CATTLE
PCR TESTS (Antigen detection)



Bovine Coronavirus realtime PCR

Material: feces, rectal swabs, small intestine

Campylobacter differentiation PCR

Detection of *Campylobacter* spec., *C. coli* and *C. jejuni*

Material: feces, rectal swabs, small intestine

Chlamydia realtime PCR

Material: Genital swabs, conjunctival swabs, placenta, fetal liver, lung, spleen, genital organs

Clostridioides (Clostridium) difficile multiplex PCR

Detection of genes coding for toxin B and the binary toxin B

Material: feces, rectal swabs

Leptospira realtime PCR

Detection incl. differentiation of pathogenic *Leptospira*
(subclades P1 and P2)

Material: Genital swabs, urine, sperm, kidney, fetal tissues (kidney, liver, lung), placenta, genital organs

Mycobacterium avium subsp. paratuberculosis PCR

Material: feces, rectal swabs, milk, mesenteric lymph nodes, small intestine

Mycoplasma (*Mycoplasma*) *bovis* PCR

Material: synovia, milk, (sperm, BALF, lung)

Rotavirus Group A and C realtime PCR

Material: feces, rectal swabs, small intestine

CATTLE
MULTIPLEX PCR TESTS (Antigen detection)



Multiplex-PCR „Respiratory pathogens Cattle”

Material: Nasal swabs, BALF, TBS, Tracheal swabs, lung

Mycoplasma (*Mycoplasma*) *bovis*

Histophilus somni

Pasteurella multocida

Mannheimia haemolytica

Bovine Coronavirus

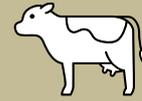
BRSV

PI3 Virus

CATTLE

SCREENINGS

Combinations of different examination methods



Mycobacterium avium* subsp. *paratuberculosis ELISA and PCR

Material: Milk on request

Mycoplasma* *bovis ELISA and PCR

Material: Milk on request

Screening „Diarrhoea calf“

Material: feces, rectal swabs, small/large intestine

Rotavirus Group A and C PCR

Bovine Coronavirus PCR

Detection of Parasites (Flotation)

Cryptosporidia special staining

Bacteriological culture for *E. coli* and *C. perfringens*

Optional, if positive:

typing *E. coli* (Multiplex-PCR for detection of 21 virulence associated factors)

typing *C. perfringens* (Multiplex PCR & Immunoblot)

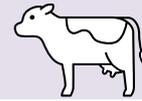
Resistance / Susceptibility testing of bacterial isolates

Agar diffusion test

Microdilution technique

(determination of the minimal inhibitory concentrations (MICs))

Microdilution technique anaerobes



Histological examination (standard staining)

Special stainings (Ziehl-Neelsen, Gram, etc.)

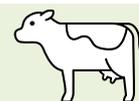
Information for sampling technique

Tissue samples for histological and immunohistological examinations should be sent in at least **10 times the volume** of 4 or 10% fixed formalin and should not be thicker than **1 cm** in at least one dimension to ensure a sufficiently rapid penetration with the fixing solution.

A cover letter with a clinical preliminary report is essential for the interpretation of the histological findings and the limitation of the deducible etiological differential diagnoses.

Immunohistological examinations for the specific detection of certain pathogens in the tissue are only useful, if a previous histological examination revealed morphological changes, which justify the suspicion of infection with the pathogen in question. Is the meaning, which a pathogen has played in a specific disease process, not of interest, but just the question, whether or not an animal was infected with the pathogen, a PCR analysis of unfixed tissue is preferable, due to its higher sensitivity.

Necropsy of ruminants is not possible due to animal health legislation regulations.



Examination by bacteriological culture, basic

Examination by bacteriological culture incl. anaerobes

using enrichment and selective culture media

Examination by bacteriological culture only *Salmonella* according

DIN EN ISO 6579 1 using enrichment and selective culture media

Note: This is no official examination according to the federal regulations against Salmonellosis in Cattle (RindSalmV)

Examination by bacteriological culture incl. *Salmonella* according

DIN EN ISO 6579 1 using enrichment and selective culture media

Note: This is no official examination according to the federal regulations against Salmonellosis in Cattle (RindSalmV)

Examination by bacteriological culture: *E.coli*, *Cl. perfringens* and *Salmonella* according DIN EN ISO 6579 1 using enrichment and selective culture media

Note: This is no official examination according to the federal regulations against Salmonellosis in Cattle (RindSalmV)

Examination by bacteriological culture incl. *Mycoplasma*

using selective culture media and enrichment culture

Note: special transport medium can be made available

Additional selective culture medium

Examination by bacteriological culture on milk (single milking)

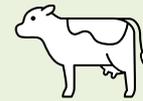
incl. yeast and molds by using selective cultural media

Please note that the identification of pathogenic bacteria species may incur further costs for diagnostic verification, e.g. for species identification by PCR or 16S rRNA typing (PCR and sequencing° (by external lab) including sequence analysis).

Most suitable sample material for cultural detection of bacterial pathogens of:

| | |
|-------------------------------|--|
| Arthritis | swab from joint capsule or joint cartilage, (synovia) |
| Respiratory infections | lung, lung and bronchial swab, bronchoalveolar lavage fluid (BALF) |
| Endometritis | cervical swab |
| Enteritis, Diarrhoea | feces, fecal swab, rectal swab, small and large intestine |
| Meningitis | meningeal swab, CSF (liquor) |
| Rhinitis | nasal swab |
| Sepsis | kidney, spleen |

Note: **swabs with medium** are the most suitable for cultural examination.



Identification of bacteria isolated at IVD as part of the cultural examination

Identification of species by PCR:

Clostridioides difficile

Fusobacterium necrophorum

Histophilus somni

Mannheimia haemolytica

Mycoplasma mycoides (*Mycoplasma*) *bovis*

Mycoplasma mycoides (*Mycoplasma*) *bovirhinis*

Pasteurella multocida

Staphylococcus chromogenes

Streptococcus parauberis

Streptococcus uberis

16S rRNA sequencing° (for identification of any bacterial species)

ITS sequencing° (for identification of further mycological species)

Cultural biochemical or serological differentiation

Resistance / Susceptibility testing of bacterial isolates

Agar diffusion test

Microdilution technique

(determination of the minimal inhibitory concentrations (MICs))

Microdilution technique anaerobes

Custodial storage of bacterial isolates / mycoplasma

Delivery of characterised bacterial field isolates

e. g. *Mycoplasma*, *Mannheimia*, *Pasteurella* and others

microbiological disinfection check°

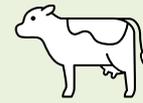
Total bacterial count and viable count *E. coli*, *C. perfringens* and mold fungi

Note: pre-register one week in advance, please

microbiological disinfection check°

only total bacterial count

Note: pre-register one week in advance, please



Identification / differentiation of submitted bacterial isolates

by PCR

Clostridioides difficile

Fusobacterium necrophorum

Histophilus somni

Mannheimia haemolytica

Mycoplasma (*Mycoplasma*) *bovis*

Mycoplasma (*Mycoplasma*) *bovirhinis*

Pasteurella multocida

Staphylococcus chromogenes

Streptococcus parauberis

Streptococcus uberis

Additional subculture for submitted bacterial isolates

for further analysis

Identification of further bacterial species by 16S rRNA typing°
ITS sequencing° (for identification of further mycological species)

Typing of bacterial isolates

***Clostridioides (Clostridium) difficile* multiplex PCR**

Detection of the genes coding for toxin A and B and the
binary toxins A and B

Clostridium perfringens

Determination of types A to F with/without β 2 toxin gene/enterotoxin gene
by PCR and detection of α and β 2 toxin production by immunoblot
only by PCR

***Escherichia coli* “Virulence associated factors” multiplex PCR**

Detection of virulence-associated factor genes (fimbriae, adhesins,
toxins, and other factors)

MRSA Methicillin resistant *Staphylococcus aureus*

Detection of *mecA* gene by PCR and of expression of penicillin-binding
protein 2 (PBP2) by agglutination

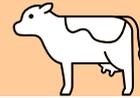
***Pasteurella multocida* capsule typing PCR**

Detection of capsular type A, B, D, E and F as well as haemorrhagic
septicaemic (HS) capsular type B

***Salmonella* Serotyping** (according to the Kauffmann-White scheme)

Classification into serogroups A-E and F-67 by agglutination

***Staphylococcus aureus*. Methicillin resistant s. MRSA**



Detection of parasite stages in feces Sedimentation Flotation
Helminth eggs, protozoan oocysts and cysts
Material: at least 10 g feces

Nematode larvae Baermann-Wetzel migration technique
of lung worms (*Dictyocaulus filaria*) and *Protostrongylidae*
Material: at least 10 g feces

Cryptosporidia Staining according to Heine
Semi-quantitative direct detection of *Cryptosporidium* oocysts
Material: 2-5 g feces (if possible, fresh and cooled, but not frozen!) or
fecal smear

Liver and rumen fluke eggs (*Fasciola*, *Dicrocoelium*)
Detection of eggs by sedimentation
Material: at least 10 g feces

Detection of skin parasites:

Mites Microscopy, KOH procedure
Sarcoptes, *Chorioptes*, *Psoroptes* mange, etc.
Material: deep skin scrapings

***Mallophaga*, lice** Microscopy

Material: 3 transparent adhesive film strips per animal on microscope slides,
combing and skin material



Avian Metapneumovirus° PCR

Subtypes A and B

Material: Pharyngeal swabs, lung, air sac/ bronchus swab

***Histomonas meleagridis*° PCR**

Material: feces, cloacal swabs, caecum

Infectious Bronchitis Virus (IBV)° PCR

Material: Pharyngeal swabs, lung, bronchus swab

Marek Disease Virus (MD)° PCR

Material: feces, cloacal swabs, feather follicles, liver, spleen, kidney, ovar, tumor tissue

***Mycoplasma*° PCR**

Mycoplasma (*Mycoplasma*) *gallisepticum* and

Mycoplasma (*Mycoplasma*) *synoviae*

Material: Pharyngeal swabs, lung, air sac/ bronchus swab

***Salmonella species* realtime PCR after cultural enrichment**

Material: feces, cloacal swabs, feather follicles, liver, spleen, kidney, ovar, tumor tissue, sock swabs, environmental swabs, bacterial isolates
on request

***Salmonella-Enteritidis-Typhimurium* realtime multiplex PCR**
Detection of *Salmonella* and identification of serovar Enteritidis and Typhimurium

Material: feces, cloacal swabs, feather follicles, liver, spleen, kidney, ovar, tumor tissue, sock swabs, environmental swabs, bacterial isolates
on request



Examination by bacteriological culture, basic

Examination by bacteriological culture incl. anaerobes

using enrichment and selective culture media

Examination by bacteriological culture incl. *Salmonella* of feces

according to DIN EN ISO 6579-1 using enrichment and selective culture media

Examination by bacteriological culture ONLY for *Salmonella* of feces and Sock swabs

according to DIN EN ISO 6579-1 using enrichment and selective culture media

***Salmonella* Serotyping** (according to the Kauffmann-White scheme)

Classification into serogroups A-E and F-67 by agglutination

***Salmonella* Serotyping of TOP5 Serovars** (according to the Kauffmann-White- Le-Minor scheme)

Detection *S. Enteritidis*, *S. Typhimurium*, *S. Infantis*, *S. Virchow*, *S. Hadar* by agglutination

Differentiation of *Salmonella* spp. vaccine and field strains

Differentiation of the vaccine strains of Cevac Salmovac

Examination by bacteriological culture incl.

Yersinia pseudotuberculosis

using selective culture media - on request

Examination by bacteriological culture incl. *Mycoplasma*

using selective culture media and enrichment culture

Additional selective culture medium

Please note that the identification of pathogenic bacteria species may incur further costs for diagnostic verification, e.g. for species identification by PCR or 16S rRNA typing (PCR and sequencing° (by external lab) including sequence analysis).

Identification of further bacterial species by 16S rRNA sequencing°

ITS sequencing° (for identification of further mycological species)

Cultural biochemic and serologic differentiation

Custodial storage of bacterial isolates / mycoplasma



microbiological disinfection check°

Total bacterial count and viable count *E. coli*, *C. perfringens* and mold fungi

Note: pre-register one week in advance, please

microbiological disinfection check°

only total bacterial count

Note: pre-register one week in advance, please

Resistance / Susceptibility testing of bacterial isolates

Microdilution technique (determination of the minimal inhibitory concentrations (MICs))

Microdilution technique anaerobes

More diagnostic using selective media are possible on request:

- *Mycobacteria*
- skin fungi
- differentiation between *Aspergillus flavus* and *A. niger*

Identification of bacterial isolates (prices incl. culture)

Avibacterium gallinarum / *paragallinarum*

Bordetella avium

Clostridium colinum

Clostridium perfringens

Enterococcus cecorum

Enterococcus faecalis

Enterococcus hirae

Erysipelothrix rhusiopathiae (Erysipelas Erreger)

Escherichia coli

Gallibacterium anatis

Mycoplasma (*Mycoplasma*) *gallisepticum* and

Mycoplasma (*Mycoplasma*) *synoviae*

Ornithobacterium rhinotracheale (ORT)

Pasteurella multocida

Riemerella anatipestifer

Salmonella species

Staphylococcus aureus. MRSA diagnostic is possible s. MRSA

Streptococcus zooepidemicus. and further *Streptococci*

Yersinia pseudotuberculosis



Most suitable sample materials for cultural detection of bacterial pathogens of:

| | |
|-----------------------------------|--|
| Arthritis | Swabs of joint capsule or cartilage, (synovia) |
| Respiratory infections | Lung, air sac, lung/bronchus swabs, pharyngeal swabs |
| Skin lesions | dermal swabs, abscess content or swabs from abscesses |
| Inflammation of ovary duct | cloacal swabs |
| Enteritis, diarrhoea | feces, cloacal swabs, rectal swabs, small /large intestine |
| Meningitis | meningeal swabs, CSF (Liquor) |
| Sepsis | kidney, spleen |
| Serositis | serosal swabs |

Note: **swabs with medium** are the most suitable for cultural examination.



Typing of bacterial isolates

Clostridium perfringens

Determination of types A to F with/without β 2 toxin gene/enterotoxin gene by PCR and detection of α and β 2 toxin production by immunoblot
nur PCR

MRSA Methicillin resistant *Staphylococcus aureus*

Detection of the mecA-Gen by PCR and expression of the Penicillin binding Protein 2 (PBP2) by agglutination

***Pasteurella multocida* capsule typing PCR**

Detection of capsular type A, B, D, E and F as well as haemorrhagic septicaemic (HS) capsular type B

Additional subculture of submitted bacterial isolates

for further analysis

POULTRY PARASITOLOGY°



Information concerning the parasitological examination of feces:

For the parasitological examination fresh fecal samples or pool samples from 3 consecutive days are most useful.

Detection of parasite stages in feces

Helminth eggs, protozoan oocysts and cysts

incl. *Eimeria* spp., *Ascaridia galli*, *Capillaria contorta*, *C. annulate*, *C. obsignata*, *C. caudinflata*, *Syngamus trachea*, *Heterakis gallinarum*, *Davainea proglottina*, *Railletina echinobothridia* etc.

Sedimentation-Flotation (qualitative, semi-quantitative)

Material: at least 10 g feces, sock swabs

Modified McMaster procedure (quantitative)

Determination of eggs/ oocyst per gram feces

threshold: 50 EPG/OPG

Material: at least 10 g feces, sock swabs

Mites

***Dermanyssus gallinae* (Poultry Red Mites), Air sac mites**

Makroskopy, Mikroskopy

Material: carcasses

Scaly leg mites (*Knemidocoptes mutans*) Mikroskopy, KOH-method

Material: skin from altered legs

Feather mites, Fleas, *Mallophagida*, Lice Mikroskopy

Material: 3 transparent adhesive film strips per animal on microscope slides, skin material, carcasses

POULTRY PATHOLOGY



NECROPSY

incl. final disposal of animal carcasses
per animal

HISTOLOGY



Histological examination

per sample



Information for sampling technique

Tissue samples for histological examinations should be sent in at least **10 times the volume** of 4 or 10% fixed formalin and should not be thicker than **1 cm** in at least one dimension to ensure a sufficiently rapid penetration with the fixing solution.

A cover letter with a clinical preliminary report is essential for the interpretation of the histological findings and the limitation of the deducible etiological differential diagnoses.

LIST OF ABBREVIATIONS

| | |
|-------|--|
| BALF | broncho-alveolar lavage fluid |
| CNS | central nervous system |
| CSF | cerebrospinal fluid, <i>Liquor cerebrospinalis</i> |
| EDTA | ethylene diamine tetraacetic acid sodium salt |
| ELISA | enzyme-linked immunosorbent assay |
| HI | hemagglutination inhibition test |
| IFAT | immunofluorescence antibody test |
| IHA | indirect hemagglutination assay |
| IHC | immunohistochemistry |
| CFT | complement fixation test |
| MAT | microscopic agglutination test |
| PCR | polymerase chain reaction |
| qPCR | quantitative polymerase chain reaction |
| RBT | Rose Bengal test |
| rtPCR | realtime polymerase chain reaction |
| SLA | serum agglutination test |
| TBS | tracheobronchial secretion |

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